

SUPPLEMENTAL INFORMATION

“Calcium release channel RyR2 regulates insulin release and glucose homeostasis”

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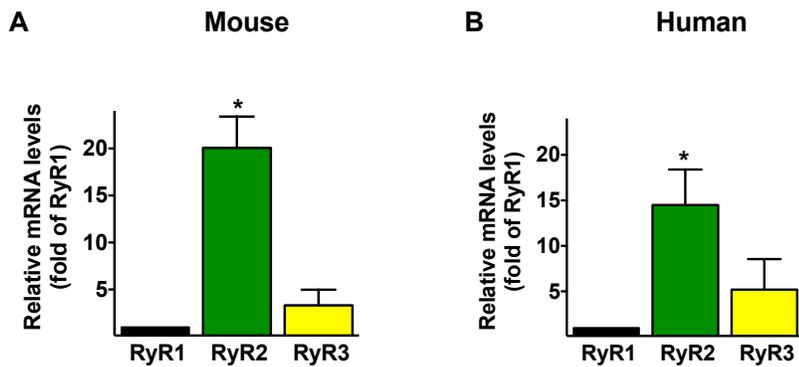
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Figure S1. RyR2 is the major form of RyR expressed in murine and human pancreatic islets. The expression of RyR1, RyR2, and RyR3 in murine (A) and human (B) islets were determined by real-time RT-qPCR analysis of total RNA, using β actin as internal standard. Primer sequences are reported in Supplementary Table 3. Each bar represents mean \pm s.e.m. of five independent experiments in each of which reactions were performed in triplicate. * P <0.05 vs RyR1; ANOVA.

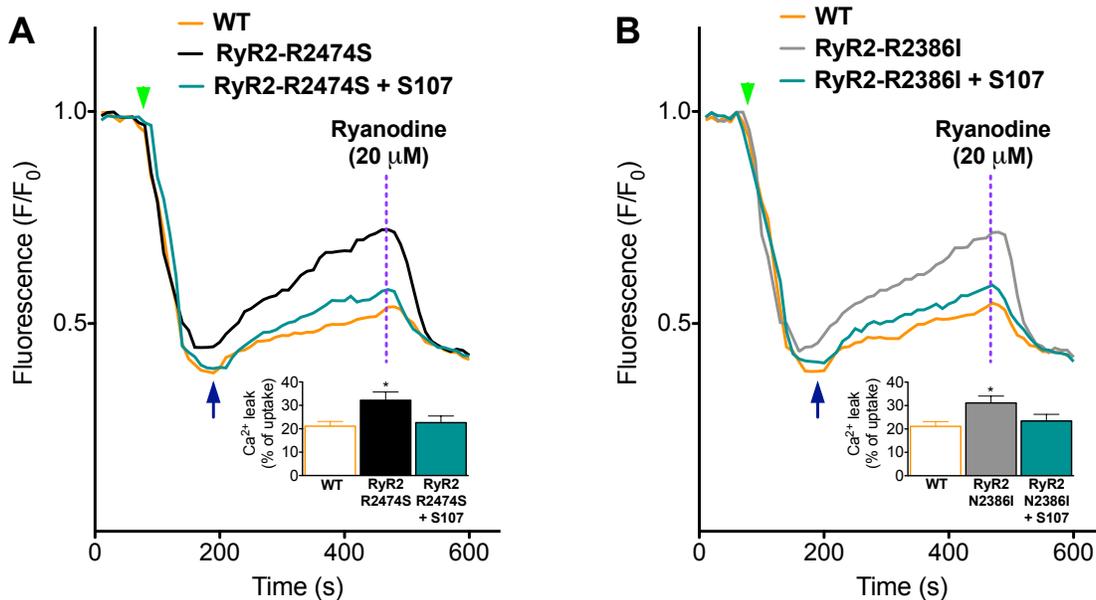


Figure S2. Increased ER Ca²⁺ leak in pancreatic islet microsomes from CPVT mice is reduced by oral treatment with S107 to fix the RyR leak. Representative traces of ER Ca²⁺ leak from pancreatic islet microsomes isolated from RyR2-R2474S (A) and RyR2-N2386I (B) mice treated with vehicle or S107 for four weeks (50 mg/kg/d in drinking water). The green arrowhead represents the addition of ATP; the blue arrow is thapsigargin (see Methods for details). The quantification of the Ca²⁺ leak (from triplicate experiments) is shown in the bar graphs in the insets. * $P < 0.05$ vs WT, ANOVA. Dashed line indicates the addition of ryanodine which identifies the source of the leak as the RyR channel.

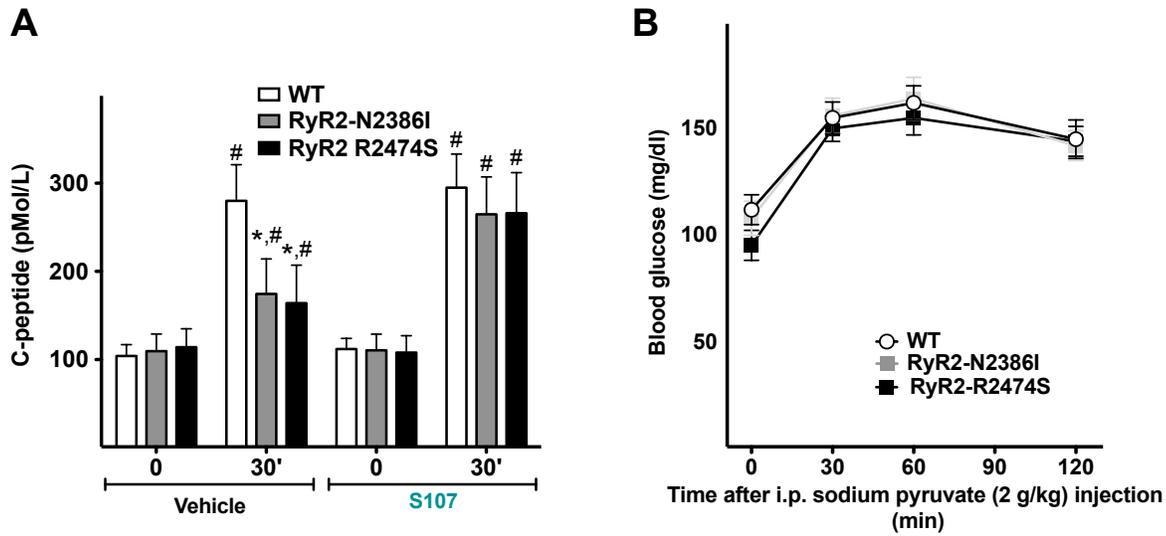


Figure S3. Evaluation of C-peptide secretion and gluconeogenesis in CPVT mice. *In vivo* determination of C-peptide release in basal conditions and 30' after an intraperitoneal (i.p.) injection (2 g/Kg) of glucose (**A**) and blood glucose levels following i.p. sodium pyruvate challenge in WT, RyR2-R2474S, and RyR2-N2386I mice (**B**). Data are expressed as mean±s.e.m. n=8-10/group. *: $P < 0.05$ vs WT; #: $P < 0.05$ vs baseline; ANOVA, Tukey-Kramer *post hoc* test.

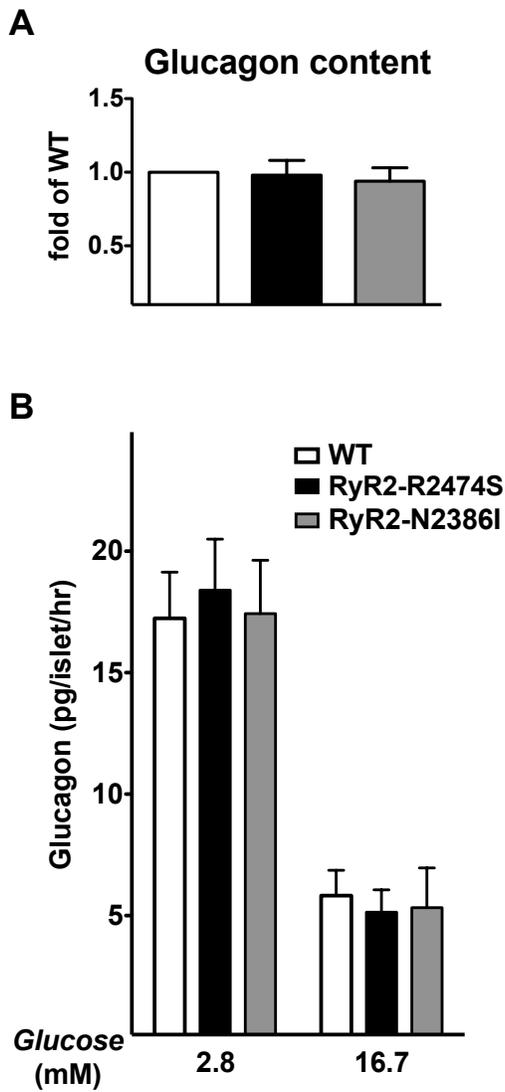


Figure S4. Glucagon content and secretion in WT and CPVT islets. Glucagon content (**A**) and secretion (**B**) in intact islets isolated from WT, RyR2-R2474S, and RyR2-N2386I mice. Data are shown as mean \pm s.e.m. from at least 6 different animals/group.

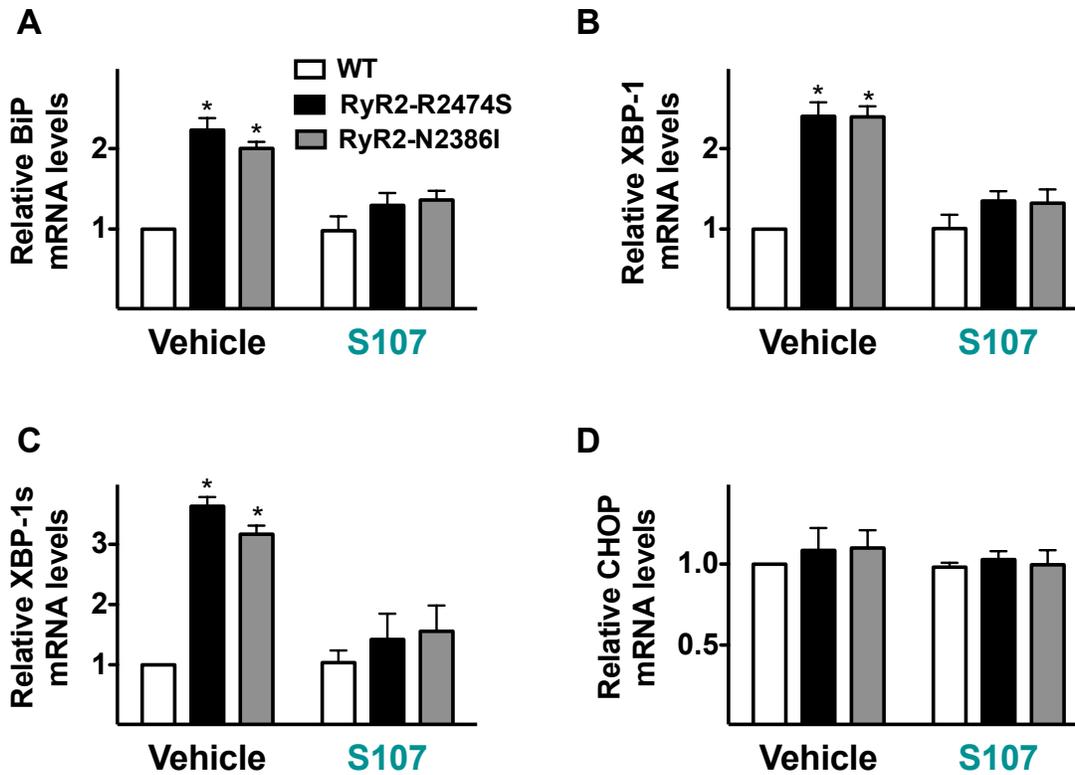


Figure S5. Activation of ER stress response in CPVT islets. The mRNA expression of BiP (A), total (unspliced) and spliced XBP-1 (B, C), and CHOP (D) was determined in murine WT and CPVT islets by real-time RT-qPCR analysis of total RNA, using β actin as internal standard. Primer sequences are reported in Supplementary Table 3. Each bar represents mean \pm s.e.m. of four independent experiments in each of which reactions were performed in triplicate. *: $P < 0.05$ vs WT; ANOVA, Tukey-Kramer *post hoc* test.

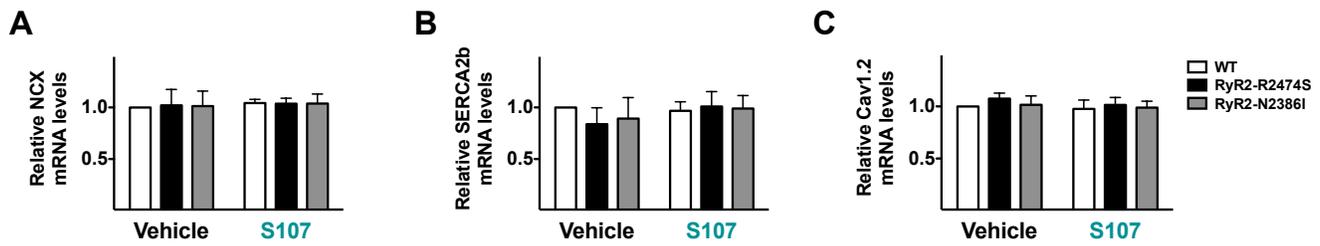


Figure S6. Expression of Ca^{2+} handling proteins in pancreatic islets of Langerhans isolated from CPVT mice. The mRNA expression of NCX (A), SERCA2b (B), and L-type Cav1.2 Ca^{2+} channel (C) was determined in murine WT and CPVT islets by real-time RT-qPCR analysis of total RNA, using β actin as internal standard. Primer sequences are reported in Supplementary Table 3. Each bar represents mean \pm s.e.m. of three independent experiments in each of which reactions were performed in triplicate.

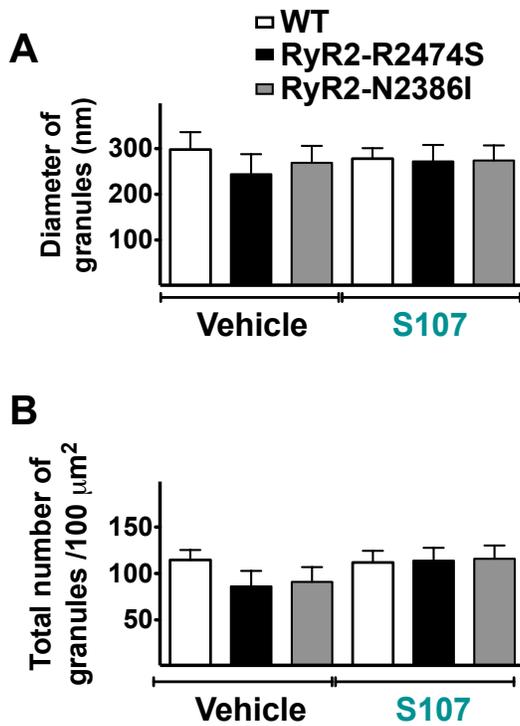


Figure S7. Quantification of the ultrastructural analyses of insulin granules in pancreatic β -cells.

Diameter (**A**) and total number (**B**) of insulin secretory granules at the electron microscope analysis of murine WT and CPVT β -cells (representative pictures are shown in **Fig. 3A**). All data are shown as mean \pm s.e.m.

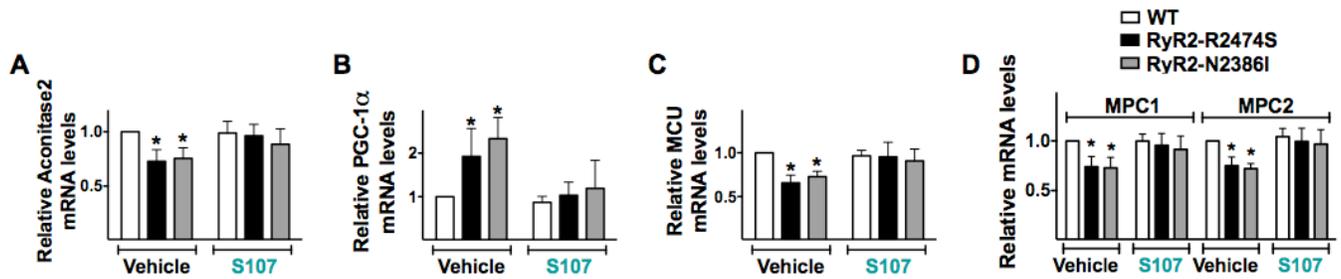


Figure S8. Expression of markers of mitochondrial dysfunction in pancreatic islets of Langerhans isolated from CPVT mice. The mRNA expression of aconitase2 (**A**), PGC-1 α (**B**), MCU (**C**), and MPC1/2 (**D**) was determined in murine WT and CPVT islets by real-time RT-qPCR analysis of total RNA, using β actin as internal standard. Primer sequences are reported in Supplementary Table 3. Each bar represents mean \pm s.e.m. of three independent experiments in each of which reactions were performed in triplicate. *: $P < 0.05$ vs WT; ANOVA, Tukey-Kramer *post hoc* test.

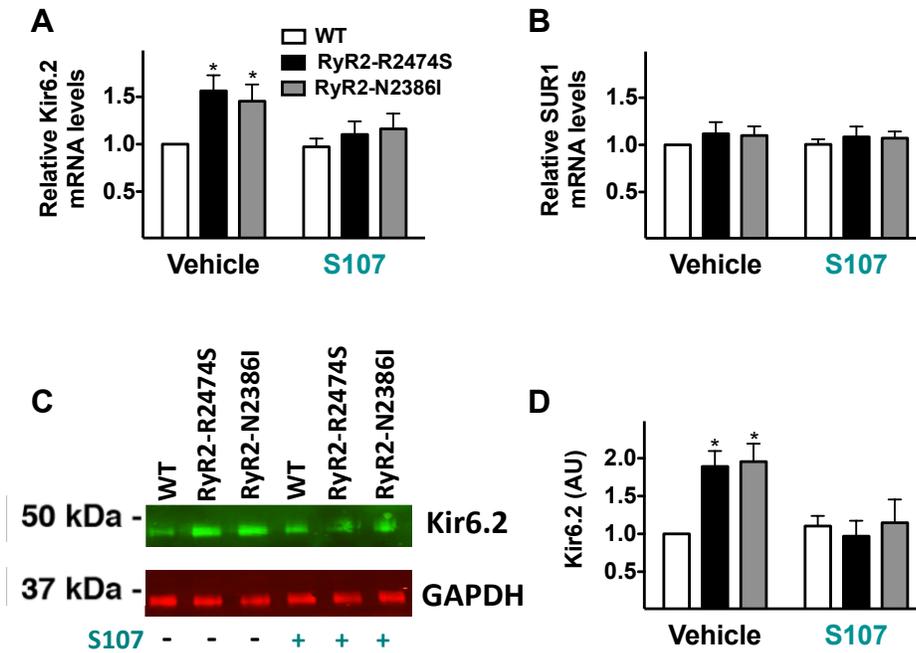


Figure S9. Increased K_{ATP} channel expression in pancreatic islets from CPVT mice. (A-B) Kir6.2 (A) and SUR1 (B) mRNA levels determined in murine WT and CPVT islets by real-time RT-qPCR analysis of total RNA, using β actin as internal standard. Primer sequences are reported in Supplementary Table 3. Mice were treated with vehicle or S107. Each bar represents mean \pm s.e.m. of three independent experiments in each of which reactions were performed in triplicate. (C) Representative immunoblots of pancreatic islets from the indicated groups of mice treated or not with S107 (50 mg/kg/d, 4 weeks) and (D) relative quantification (triplicate experiments); AU: arbitrary units; *: $P < 0.05$ vs WT vehicle; ANOVA, Tukey-Kramer *post hoc* test.

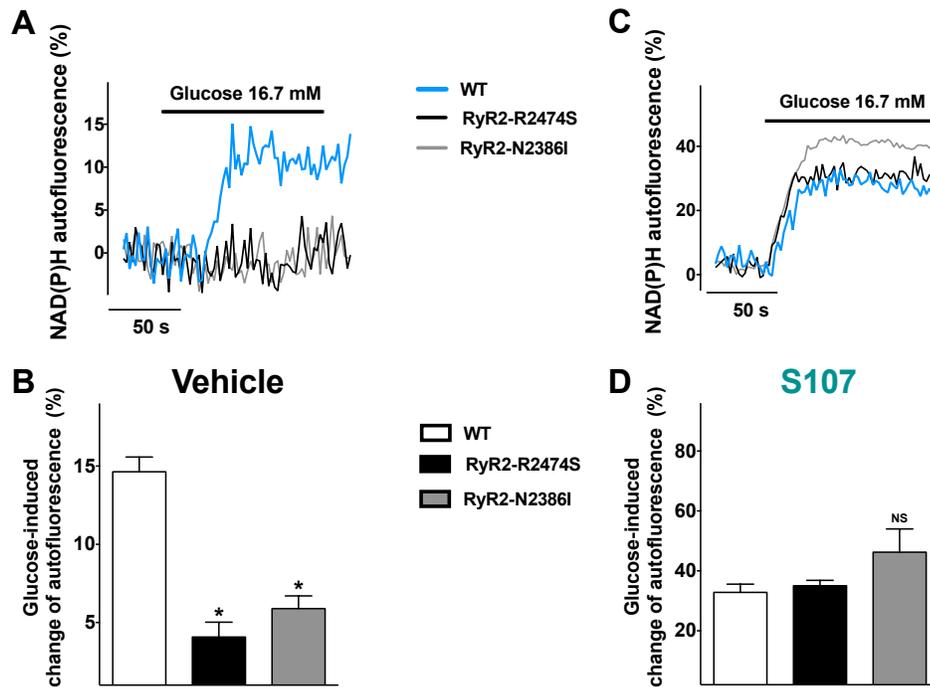


Figure S10. Blunted glucose-induced NAD(P)H autofluorescence in CPVT mice. (A-D) Evaluation of NAD(P)H autofluorescence in response to glucose dispersed cells from islets isolated from WT and CPVT mice treated with vehicle (A, B) or with S107 (50 mg/kg/d, 4 weeks, C, D). In A and C representative traces are depicted. 7-9 mice per group were used. Data are shown as mean±s.e.m. *: $P < 0.05$ vs WT; ANOVA, Tukey-Kramer *post hoc* test. Additional details are given in Methods.

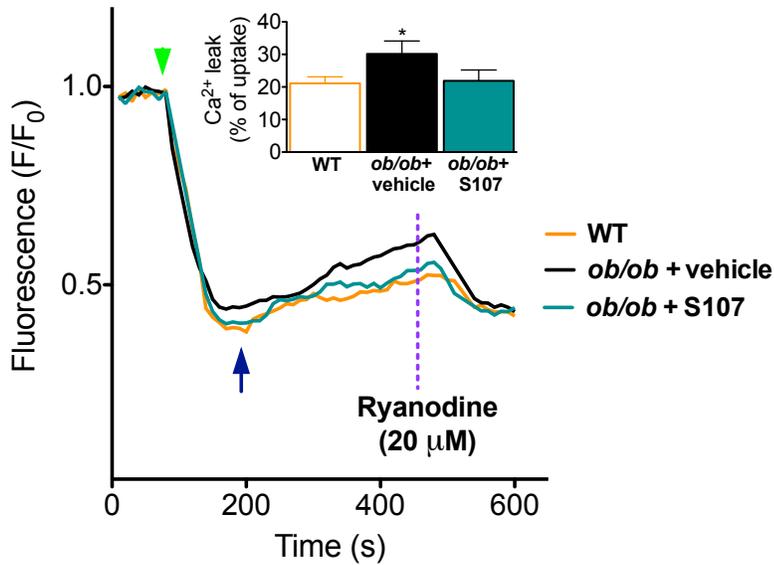


Figure S11. Increased ER Ca²⁺ leak in pancreatic islet microsomes from *ob/ob* mice is reduced by oral treatment with S107 to fix the RyR leak. Representative traces of ER Ca²⁺ leak from pancreatic islet microsomes isolated from *ob/ob* mice treated with vehicle or S107 for four weeks (50 mg/kg/d in drinking water). The green arrowhead represents the addition of ATP; the blue arrow is thapsigargin (see Methods for details). The bar graph in the inset is the quantification of Ca²⁺ leak (from triplicate experiments). **P* < 0.05 vs WT, ANOVA. Dashed line indicates the addition of ryanodine.

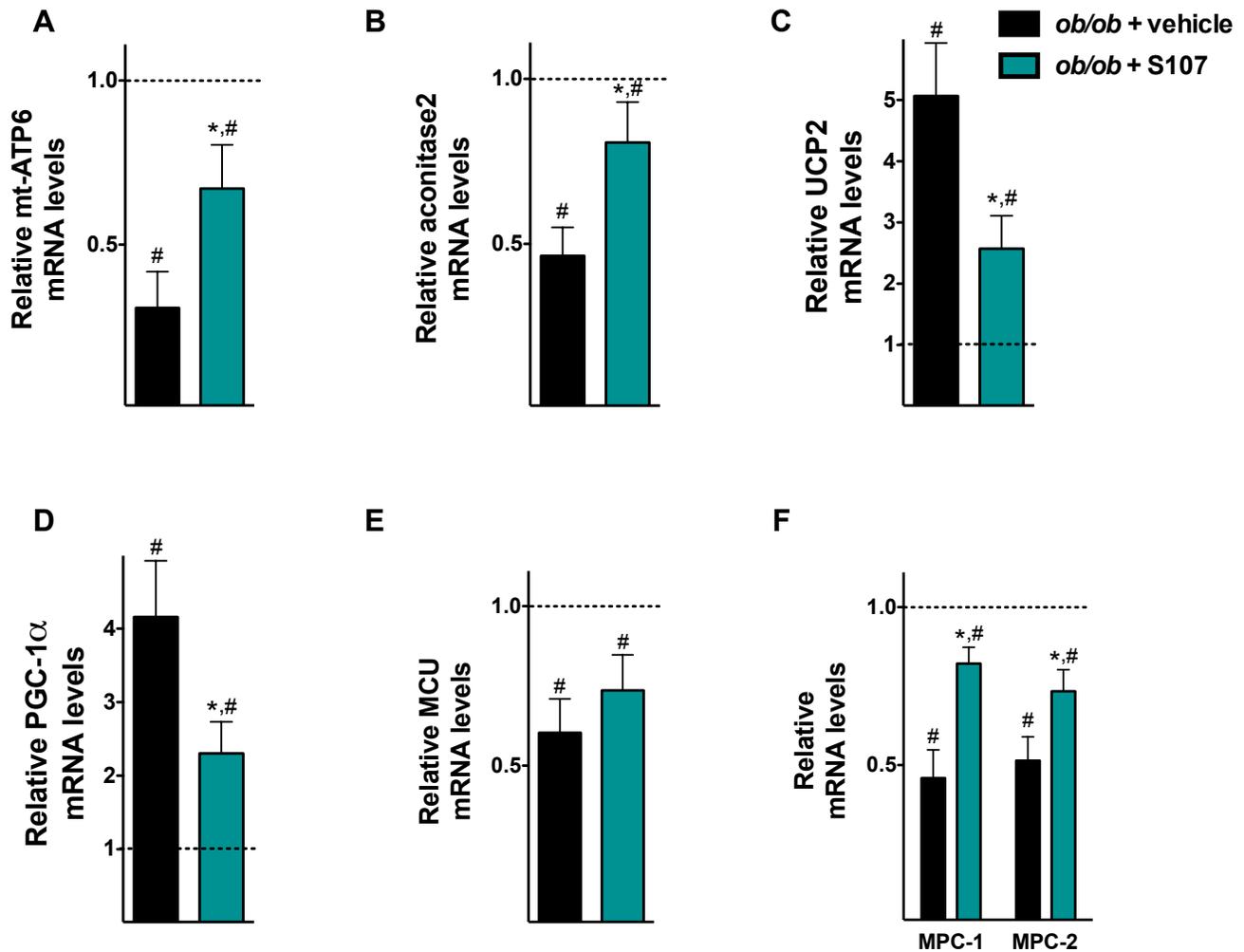


Figure S12. Effects of chronic S107 treatment on markers of mitochondrial dysfunction in pancreatic islets of Langerhans isolated from *ob/ob* mice. The mRNA expression of mt-ATP6 (A), aconitase 2 (B), UCP2 (C), PGC-1 α (D), MCU (E), and MPC1/2 (F) in *ob/ob* mice treated or not with S107 (50 mg/kg/d, 4 weeks) was determined by real-time RT-qPCR analysis of total RNA, relative to untreated WT mice (horizontal dashed lines), using β actin as internal standard. 7-9 animals per group were used. Primer sequences are reported in Supplementary Table 3. Each bar represents mean \pm s.e.m. of three independent experiments, in each of which reactions were performed in triplicate. *: $P < 0.05$ vs *ob/ob*+vehicle; #: $P < 0.05$ vs WT; 2 tailed Student's *t* test.

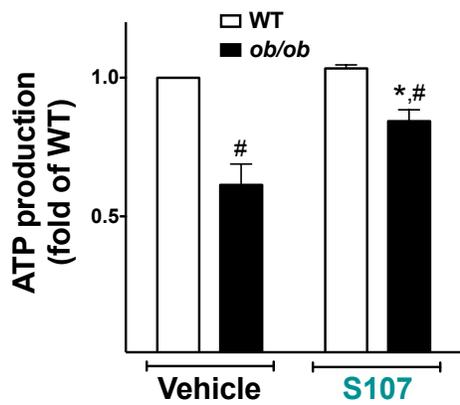


Figure S13. Reduced ATP production in *ob/ob* islets is ameliorated by S107. Chronic S107 treatment of *ob/ob* mice (50 mg/kg/d, 4 weeks) improved ATP production in response to methyl pyruvate (10 mM) in isolated islets of Langerhans. Each bars represents mean±s.e.m. from at least 6-8 animals/group.

*: $P < 0.05$ vs *ob/ob*+vehicle; #: $P < 0.05$ vs WT; 2 tailed Student's *t* test.

Supplementary Table 1. Baseline clinical characteristics of control and CPVT subjects.

	Control (n=27)	CPVT (n=27)
Age (years)	27.2±4.1	26.9±3.7
Male (%)	59.25	62.9
BMI (kg/m²)	23.6±2.4	21.7±2.9
Fasting blood glucose (mg/dl)	94.4±8.1	92.2±7.4
Fasting serum insulin (μU/ml)	7.76±2.11	7.93±2.29
Waist circumference (cm)	41.6±5.2	42.1±5.4
Waist-to-hip ratio	0.93±0.08	0.90±0.11
Systolic blood pressure (mmHg)	126.6±9.2	128.3±8.8
Diastolic blood pressure (mmHg)	75.8±3.6	72.4±3.8
Triglycerides (mg/dl)	121.8±32.5	118.1±29.8
Total cholesterol (mg/dl)	184.4±38.4	186.1±41.2
HDL cholesterol (mg/dl)	47.6±12.7	48.2±14.1
LDL cholesterol (mg/dl)	112.1±27.7	114.3±28.1
Creatinine (mg/dl)	0.86±0.18	0.91±0.24
Blood urea nitrogen (mg/dl)	11.4±3.1	10.8±3.2
Current cigarette smokers (%)	18.75	12.5
Familial sudden cardiac death (%)	-	40.7
Implanted cardioverter defibrillator (%)	-	81.48

Clinical characteristics of the subjects, including body mass index (BMI) high-density (HDL) and low-density (LDL) lipoprotein cholesterol. CPVT stand for Catecholaminergic Polymorphic Ventricular Tachycardia. Values of serum insulin were available in 19/27 individuals per group. Data are expressed as mean±s.e.m. or percentage.

Supplementary Table 2. RyR2 mutations in CPVT patients.

# Patient	RyR2 Mutation	Blood glucose (mg/dl) 2h after OGTT
#1	P2404T	187
#2	K4392R	147
#3	L2534V	192
#4	T2510A	188
#5	R169Q	166
#6	R2401L	188
#7	R4959Q	131
#8	R2474S	199
#9	V2475F	191
#10	P2328S	196
#11	R169Q	172
#12	R2401H	174
#13	Q4201R	133
#14	H4108N	122
#15	I4587V	135
#16	E2296Q	191
#17	R1051P	189
#18	H4762P	194
#19	N2386I	198
#20	N1551S	144
#21	G4671R	189
#22	A2387P	185
#23	S4565R	145
#24	V2113M	188
#25	E1724K	186
#26	F2307L	193
#27	N1551S	192

Supplementary Table 3. Primer sequences.

Gene	Forward 5'-3'	Reverse 5'-3'	Product size (bp)
Human RyR1	TTCCCAAGATGGTGACAAG	CTCCAGCAGGTAGCTCAGGT	105
Human RyR2	CCTTGCCTGAGTGCAGTTG	TTGAGGTATCAACAGGTTGTGG	130
Human RyR3	ATCAACACGCCATCTTTTCC	TTGTCCAAACCCAGGAGTTC	119
Human β-Actin	CTCTTCCAGCCTTCCTCCT	AGCACTGTGTTGGCGTACAG	116
Mouse RyR1	CCTTGCCTGAGTGCAGTTG	TTGAGGTATCAACAGGTTGTGG	130
Mouse RyR2	TTCCCAAGATGGTGACAAG	CTCCAGCAGGTAGCTCAGGT	105
Mouse RyR3	AGATCGAGCTGCTGAAGGAA	CCTTCCAGGAGGGACAGAAG	73
Mouse Kir6.2	GTAGGGGACCTCCGAAAGAG	CTGGTGGAGAGGCACAACCTT	102
Mouse SUR1	GAACAGGCAAGACCAAGAGC	CAGAAGGCCATCTCTTGAGG	106
Mouse CaV1.2	CAGCCCAGAAAAGAAACAGG	GGATTCTCCATCGGCTGTAA	100
Mouse NCX	AGATCAAGCATCTGCGTGTG	CTCCACAACCTCCAGGAGAGC	104
Mouse SERCA2b	AGGGACTGCAGTGGCTAAGA	GCCACAATGGTGGAGAAGTT	69
Mouse UCP2	AGCCTTCTGCACTCCTGTGT	TAGAAAATGGCTGGGAGACG	82
Mouse Aconitase2	CAGGTGCCAAGTATGGTGTG	GCCGATCAGAAGAACTCCAG	101
Mouse PGC1-α	CCGAGAATTCATGGAGCAAT	TTTCTGTGGGTTTGGTGTGA	129
Mouse mt-ATP6	CAACCGTCTCCATTCTTTCC	CGTCCTTTTGGTGTGTGGAT	81
Mouse MPC1	ACTTTCGCCCTCTGTTGCTA	ACATGGCATGCAAACAAAAG	89
Mouse MPC2	AATGAGGCCGCTTTACAAC	GCTAGTCCAGCACACACCAA	94
Mouse MCU	TCATGGAATCCAAGTGCAAA	AGCAAACACCCAACATCCTC	109
Mouse Bip	TGCAGCAGGACATCAAGTTC	TTTCTTCTGGGGCAAATGTC	111
Mouse unspliced XBP-1	TCAAATGTCCTTCCCCAGAG	AAAGGGAGGCTGGTAAGGAA	87
Mouse spliced XBP-1	CTGAGTCCGCAGCAGGTG	GGCAACAGTGTGAGAGTCCA	74
Mouse CHOP	CTGCACCAAGCATGAACAGT	CTACCCTCAGTCCCCTCCTC	131
Mouse β-Actin	GCTCTTTTCCAGCCTTCCTT	GCCTCAGGAGTTTGTGACC	78
Mouse H19	GTACCCACCTGTCGTCC	GTCCACGAGACCAATGACTG	207
Mouse 18S	CGCGTTTCTATTTTGTGGT	AGTCGGCATCGTTTATGGTC	219
Mouse mt12S	ACCGCGGTCATACGATTAAC	CCCAGTTTGGGTCTTAGCTG	178
Mouse mt16S	CCTTGTTCACAGAGGTTCAA	ATGCCGTATGGACCAACAAT	169

RyR: Ryanodine Receptor/ Ca^{2+} release channel; Kir: Potassium inwardly-rectifying channel; SUR: sulfonylurea receptor; Cav1.2: Ca^{2+} channel voltage-dependent, L-type, α 1C subunit; NCX: $\text{Na}^+/\text{Ca}^{2+}$ exchanger; SERCA: Sarco/Endoplasmic Reticulum Ca^{2+} -ATPase; UCP2: Uncoupling Protein 2; PGC1- α : Peroxisome proliferator-activated receptor gamma (PPAR γ)-coactivator 1-alpha; mt-ATPs: mitochondrially encoded ATP synthase 6; MPC: Mitochondrial Pyruvate Carrier; MCU: Mitochondrial Ca^{2+} Uniporter; BiP: immunoglobulin heavy-chain binding protein; XBP-1: X-box binding protein 1; CHOP: CCAAT/enhancer-binding protein homologous protein.

Supplementary Table 4. Characteristics of human pancreatic islet donors.

	Age	BMI	T2DM
#1	55	24	No
#2	41	35.5	No
#3	51	17.3	No
#4	32	28.2	Yes
#5	53	27.1	Yes
#6	42	30.2	Yes

BMI: Body-mass index; T2DM: Type 2 Diabetes Mellitus.

Supplementary Table 5. Baseline characteristics of WT and CPVT mice (4-month-old).

	WT	RyR2-R2474S	RyR2-N2386I
Body weight (g)	31.5±1.6	30.9±1.7	31.1±1.9
Food intake (g/day)	3.4±0.9	3.4±0.8	3.3±0.9
Water intake (ml/day)	5.9±0.8	6.3±1.1	6.2±1.1
Fasting blood glucose (mg/dl)	95.33±15.1	98.4±17.7	97.8±18.2
Fasting serum insulin (ng/dl)	0.46±0.06	0.41±0.07	0.43±0.07
Triglycerides (mM/l)	0.39±0.05	0.38±0.06	0.41±0.05
Free fatty acids (mM/l)	0.68±0.12	0.71±0.11	0.65±0.14
Mean arterial blood pressure (mmHg)	103.4±9.6	104.1±10.8	103.7±11.2

Data are expressed as mean±s.e.m from at least 6 mice per group.